RAJARSHI SHAHU MAHAVIDYALAYA, LATUR (Autonomous)

(BoS in ZOOLOGY)

CHOICE BASED CREDIT SYSTEM (CBCS) SEMESTER PATTERN

(W.e.f. Academic Year 2020-21)



SYLLABUS FOR B.Sc.-II EXAMINATION SUB: ZOOLOGY

RAJARSHI SHAHU MAHAVIDYALAYA, LATUR. (Autonomous)

SEMESTER PATTERN CURRICULUM UNDER CHOICE BASED CREDIT SYSTEM (CBCS) Faculty of Science

B.Sc. S.Y. Under Graduate (UG) Programmes SUBJECT: ZOOLOGY (w. e. f. June -2020)

Semester	Course Number		Paper No. and Title	Total period/week	Marks		Credits
					Internal	External	
					ESE	CA	
SEM-III	CCZ-III	А	Animal Physiology and Immunology	45 3/week	20	30	2
		В	Genetics	45 3/week	20	30	2
	CCZP-III	Α	Animal Physiology and Immunology	45 3/week	20	30	2
	CCZP-IV	В	Genetics	45 3/week	20	30	2
	SECZ-I		Clinical Haematology	3/Week	20	30	2
SEM-IV	CCZ-IV	Α	Biochemistry	45 3/week	20	30	2
		В	Molecular Biology and Genetic Engineering	45 3/week	20	30	2
	CCZP-V	Α	Biochemistry	45 3/week	20	30	2
	CCZP-IV	В	Molecular Biology and Genetic Engineering	45 3/week	20	30	2
	SECZ-II		Microtechniques	3/Week	20	30	2

B. Sc. II ZOOLOGY (SEM-III)

Course Code: U-ZOO-382

Course: Animal Physiology and Immunology (CCZ-III-A)

Credits: 02 Marks: 50 Lectures: 45

Learning Objectives

- To make the students to understand the physiological process of animals
- To understand the importance of physiology and Immunology
- To understand concepts and different types of immunity.
- Tohavebasicknowledgeoffunctions of different organs and organ system
- To introduce the concepts of physiology of digestion excretion and Osmoregulation
- To introduce the concepts of physiology of respiration and circulation

Course Outcome

- Learners would understand the different physiological process of animals
- Learners would be able to correlate the habit and habitat with nutritional, excretory and osmoregulatory structures
- Learners would be able describe the concepts and different types of immunity.

Unit – I (Digestion Physiology and Respiration physiology)

- Intracellular and Extracellular digestion
- Mechanical process in digestion
- Chemical process of digestion
- Absorption of food
- External and Internal respiration
- Respiratory organs of man
- Mechanism of respiration
- O2 and Co2 transportation
- Control of respiration and Respiratory quotient

Unit -II (Circulation physiology)

- Open and closed type of circulation
- Circulatory organs (Heart and Blood vessels)
- Typical pattern of circulation
- Composition and function of blood
- Level of blood Cholesterol, urea and sugar.
- Erythropoiesis and Its regulation
- Blood pressure, ECG, Heart beat and Cardiac cycle.
- Lymphatic system (Lymph, Lymph vessels and Lymph node

Unit -III (Excretion physiology and Neurophysiology)

- Mode up of Excretion
- Structure of nephron (Uriniferous tubule)
- Physiology of Urine formation Composition of urine and Osmoregulation
- Structure of Neuron
- Structure of Synapse and reflex action
- Conduction of nerve impulses and Neurotransmitter

UNIT:-IV (Muscle physiology, Reproductive Physiology and Immunology)

- Structure and types of muscles
- Ultra structure of skeletal muscle fibre
- Sliding filament theory
- Hormonal control of testicular and ovarian functions, Menstrual cycle, Estrous cycle, and Homeostasis.
- Types of Immunity ii) The immune system
- Immune response (Antigen, Antibody, Humoral and cell mediated immunity)

- A textbook of Animal Physiology K.A. Goel and K.V. Shastri (Rastogi Pub.)
- A textbook of Practical Physology V.G. Ranade (P.V.G. Prakashan Pune.)
- Clinical Pathology and Haematology Nanda Baheti (Kanhaiya Pub.)
- Comparative animal physiology C. Ladd Prosser.
- Text book of animal Physiology A.K. Berry (Emkay Pub. Delhi)
- Animal Physiology A. Mariakuttikan N. Arumugam (Saras Publication)
- Principles of animal Physiology Wood D.W
- Physiology Guyton and Hall
- Duby-Immunology (W.H. Freeman)

B. Sc. II ZOOLOGY (SEM-III)

Course Code: U-ZOO-382 Course: Genetics (CCZ-III-B)

Credits: 02 Marks: 50 Lectures: 45

Learning Objectives:

- To make the students to understand the structure and functions of gene
- To understand the importance of Genetics
- To have basic knowledge of mutation
- To understand the inheritance pattern.

Course Outcome

- Understand and apply the principles of inheritance.
- Understand the concept of multiple alleles, linkage and crossing over.
- Learner able to introduce the concept of sex determination and its types, sex
- Influenced and sex limited genes.
- Learners would understand mechanisms of sex determination.
- Learners would be able to correlate the disorders linked to a particular sexchromosome.

Unit –I (Mendelian genetics and Modifications)

- Mendelism
- Monohybrid cross and Dihybrid cross
- Interaction of gene (9:3:4, 9:7, 13:3, 15:1)
- Incomplete dominance.
- Back cross and test cross.

Unit - II (Multiple Alleles and Multiple Gene)

- Multiple alleles Eq. Coat colour in Rabbit .and ABO Blood groups in Man.
- Rh factor and Erythroblastosis foetalis in man.
- Multiple genes Eg. Skin colour in Man.
- Linkage definition, Types and significance
- Crossing over Mechanism, Factor affecting on crossing over, and Its Significance

Unit – III (Sex determination, sex linked inheritance and Gene Mutations)

- Chromosomal methods of sex determination.
- Bridge's ratio theory of genic balance.
- Sex linked inheritance in Drosophila.
- Sex linked inheritance in man colorblindness, haemophilia, Hypertrichosis and baldness.
- Chromosomal Mutations Structural and numerical mutations
- Mutagenic agents
- Sickle cell anemia.

Unit - IV (Human genetics)

- Syndromes Turner's, Klinefelter's, Down's, Cat Cry, Patau's, and Edwards.
- Inborn errors of metabolism Phenylketonuria (PKU), Alkaptonura, Albinism.
- Human Pedigree analysis with Symbols use and Eugenics and Euphenics

- Gupta, P.K. (1996) "Genetics" Rastogi Publications.
- Ranga, M.M. "Animal Biotechnology (Agrobios), Published by Agrobios (India).
- Rastogi, Sharma, V.N. and Anuradha Tandon (1993). "Concepts in Molecular Biology". WileyEastern Ltd. N. Delhi.
- Smustad, Simmons, Jenkins (1999). "Principles of Genetics" John Wiley and sons. Inc.
- Genetics P.K. Gupta (Rastogi pub. Meerut)
- Genetics Verma P.S. and Agarwal V.K. (S. Chand pub. Delhi.)
- Genetics Winchester (Oxford LBH Pub.)
- Genetics and Evolution A.P. Jha (Macmillon India)
- Concepts of genetics W.S. Clug (Pearson Education ISBN)
- Genetics Strickberger (Prentice Hall)
- Principle of genetics R.H. Tamarin (Tata Mc Graw Hill Pub. India)
- Concepts of Genetics R. L. Kotpal (Rastogi Pub.)
- Genetics and Genetic Engineering Dr. R.P. Meyyan (Saras Pub.)
- Foundations of Genetics Pai A.C. (Mc Graw Hill Pu b.)
- Molecular Genetics Gunther, S. Stent, (Macmillon)
- Principles of Genetcs Sinnott, Dunn and Dobzansky (Tata McGraw Hill Pub. Delhi).
- Genetics M.P. Arora (Himalaya).
- Genetics and Evolution N. Armugam (Saras Pub.)
- Genetic Veer Bala (Rastogi Pub.)

B. Sc. II ZOOLOGY (SEM-III)

Course Code: U-ZOO-382

Lab. Course-III: Animal Physiology and Immunology (CCZP-III-A)

Credits: 02 Marks: 50 Lectures: 45

Learning Objectives

- To make the students to understand the hematological and immunological techniques.
- To make the students to understand the hematological techniques conducted in laboratories including, complete blood count, blood grouping, blood films, and differential count.
- To make the students to understand the process of digestion by qualitative detection of digestive enzymes.
- To make the students to understand the analytical techniques know the functional status of different organ.

Course Outcome

- Learners would understand the different physiological process of animals
- Learners would be able to understand functional status of organ
- Learners would be able to understand complete blood count, blood grouping, blood films, and differential count.
- Learners would understand the respiratory status of animals
- 1. Qualitative detection of digestive enzymes (protease, Amylase and Lipase) in cockroach/ Crab.
- 2. Detection of human salivary amylase.
- 3. Estimation of oxygen consumption in fish/ Crab or any other suitable aquatic animal.
- 4. R.B.C. Counting.
- 5. W.B.C. counting.
- 6. Differential leucocyte count of blood.
- 7. Measurement of blood pressure by sphygmomanometer.
- 8. Estimation of Haemoglobin.
- 9. Estimation of urine / serum creatinine from blood
- 10. Estimation of urine / serum urea by diacetyl monoxime method
- 11. Colorimetric estimation of blood/serum cholesterol.
- 12. ESR of blood.
- 13. Determination of clotting time of blood by capillary tube method.
- 14. Estimation of glucose by Benedict quantitative method.
- 15. Determination of bilirubin in serum
- 16. Qualitative detection of Nitrogenous waste products (Ammonia, Urea and Uric acid)
- 17. Preparation of Haematin crystals.
- 18. Types of nerves Unipolar, Bipolar, Multipolar. (Chart/slides)
- 19. Antibody and antigen reaction

B. Sc.II ZOOLOGY (SEM-III)

Course Code: U-ZOO-382

Lab. Course-IV: Genetics (CCZP-IV-B)

Credits: 02 Marks: 50 Lectures: 45

Learning Objectives:

- To make the students to understand the Mendel's Laws of inheritance.
- To understand the importance modification in Mendelian laws like complementary factor to Blood group.
- To understand the problems based on sex linked inheritance.
- To understand the chromosomal abnormalities.

Course Outcome

- Learner will be able to do the problems based Mendel's laws.
- Understand and apply the concept of multiple alleles, linkage and crossing over.
- Learner able to do the problems based on sex linked inheritance and blood groups etc.
- Learners would understand mechanisms of sex determination.
- Learners would be able to understand the Human pedigree analysis and symbols.

PRACTICALS

- 1. Problems based on monohybrid and dihybrid cross (Explain with the help of plastic beads.)
- 2. Problems on modification in ratio due to interaction of genes—complementary factors, supplementary factors, inhibitory factors, duplicate genes (explain with the help of plastic beads).
- 3. Problems on blood group inheritance in man.
- 4. Problems based on sex linked inheritance
- 5. Culture of Drosophila and observation of genetic characters in Drosophila (eye & wings)
- 6. Preparation of temporary slides of salivary gland chromosomes from chironomous larva.
- 7. Study of slide of sickle cell anemia.
- 8. Study of chromosomes abnormalities in man, Down's syndrome, Klinfelter Syndrome, and Turner Syndrome with the help of Photograph / Charts / Karyotype.
- 9. Drosophila culture techniques.
- 10. Study of phenotypic characters in Drosophila (Body colour, Wing pattern and Eye colour).
- 11. Buccal smear Identification of Barr Body
- 12. Human pedigree analysis- Various symbols used and problems

B. Sc.II ZOOLOGY (SEM-III)

Course Code: U-ZOO-382

Course: Clinical Haematology (SECZ-I)

Credits: 02 Marks: 50 Lectures: 45

Learning Objective:

- To understand Perform staining and counting technique for identification of different type of blood cells.
- To understand collection methods of blood sample.
- To understand separation techniques of blood sample components.
- To understand diagnosis of different blood related diseases.

Learning Outcomes:

After completion of this course students should be able to:

- Perform staining and counting technique for identification of different type of blood cells.
- Collect blood sample by different methods.
- Separate different components of blood
- Estimation of Hb from blood samples
- Diagnosis of various blood diseases like anaemia

UNIT – I

1. The components of blood

- Plasma
- Red blood cells
- White blood cells
- Platelets

2. Collection of Blood

- Criteria for sample collection
- Collection of capillary blood (Peripheral Blood) blood by skin punctures
- Collection of venous blood by Venipuncture,
- Collection of arterial blood.

3. Practical:

- Collection Blood by Skin puncture and Venipuncture.
- Separation of Blood components Plasma, Serum and Corpuscles

UNIT - II

1. Haemoglobin

- Structure and function of Haemoglobin
- 2. Anemia Causes, Effect and Control
 - Types of anemia
 - Causes and Symptoms of anemia
 - Control measure of anemia
 - Diagnosis of anemia

3. Practical

• Estimation of Haemoglobin

UNIT – III.

- 1. Haemopoiesis, erythropoiesis and leucopoiesis
- **2. Practical:** Complete Blood Count (CBC)
 - RBC Counting

- WBC Counting
- Platelet count and Hamatocrit

UNIT - IV

- Blood Clotting,
- Mechanism of Clotting
- Extrinsic and Intrinsic Mechanism
- Blood choesterols and Urea and Creatin
- **3. Practical:** Clotting and bleeding time of blood.

REFERENCE BOOKS:

- Medical Laboratory Technology Ramnik Sood
 Medical Lab Technology Vol. I, II & III Kanai Mukherjee
- 3. Hand Book of Medical Technology Mrs. Chitra
- 4. Medical Laboratory Technology A. Ananthanarayan
- 5. Manual for Laboratory Technician of Primary Health by Minister of Health
- 6. Human Physiology Vol. I & II C. C. Chatterjee

B. Sc.II ZOOLOGY (SEM-IV)

Course Code: U-ZOO-382

Course: Biochemistry (CCZ-IV-A)

Credits: 02 Marks: 50 Lectures: 45

Learning Objectives:

- To make the students to understand chemistry of life
- To understand the importance of Biochemistry
- To have basic knowledge of metabolism
- To give learner insight into the structure of biomolecules, and their role in sustenance of life.

Course Outcome:

- The learner will realize the importance of biomolecules and their clinical significance.
- Able to demonstrate an understanding of fundamental biochemical principles, such as the structure/function of biomolecules, metabolic pathways, and the regulation of biological/biochemical processes.

Unit-I (Classification, Structural properties and biological functions)

- Introduction of Biochemistry
- Carbohydrates and lipids
- Proteins and Amino acids and peptides,

Unit-II (Classification, Structural properties and biological functions)

- Vitamins (Discovery, types and their functional Significance.)
- Fat soluble (A, D, E, K, Q and U)
- Water soluble (B-complex family and Ascorbic Acid)
- Antioxidants

Unit-III (Enzymology)

- Nomenclature of enzyme and chemistry of enzymes
- Classification, properties of enzyme
- Mechanism and factor affecting on enzyme action
- Enzyme inhibition and Biological function of enzymes
- Coenzymes

Unit-VI (Metabolism)

- Metabolism of Carbohydrates
- (EMP, HMP pathway, Krebs cycle, Glycogenosis, Glyconeogenesis, Glycogenolysis.)
- Metabolism of lipid
- (Beta oxidation pathway, Ketogenesis, ketolysis, ketosis) iii) Metabolism of protein
- (Oxidation of amino acids. Urea cycle, Transamination Deamination)
- Biochemistry of Hormones

- Lehninger Principles of Biochemistry
- Fundamentals of Biochemistry-J.L. Jain, Sunjay Jain, Nitin Jain, S. Chand & Company.
- Biomolecules-C.Kannan, MJP Publishers, Chennai-5.
- Laboratory manual in Biochemistry Jayaraman.
- Biochemical methods -S.Sadasivan and Manickam.
- Introduction to Practical Biochemistry -David T. Plumme

B. Sc. II ZOOLOGY (SEM-IV)

Course Code: U-ZOO-382

Course: Molecular Biology and Genetic Engineering (CCZ-IV-B)

Credits: 02 Marks: 50 Lectures: 45

Course Outcomes:

- To make the students to understand biological tools for research
- To understand the importance of Molecular Biology and Genetic Engineering
- To have basic knowledge of cloning and Gene Expression

Course Outcome:

- Students will able to use biological tools for research.
- Learner will able to describe the importance of Molecular Biology and Genetic Engineering.
- Learner will able to use vectors in cloning techniques and rDNA technology

UNIT- I

- Introduction to Molecular Biology
- Deoxyribonucleic acid (Structure, properties, function, and type of DNA)
- Ribonucleic acid (Structure and types)
- Replication and Genetic codes

UNIT-II

- Protein synthesis
- Gene concept and molecular structure of gene
- Types of gene and Discontinuous genes (Exons and Introns)
- Gene Expression in prokaryotes (Lac operon) eukaryotes
- One gene one enzyme hypothesis and one polypeptide hypothesis

UNIT-III

- Introduction to genetic engineering-Mendel's to Molecules
- Tools; a} Enzymes: i. lysing ii. Ligases
- iii Nucleases {Exonuceases, Endonucleases., Restrication Endonucleases enzymes}
- iv. Syntheases { DNA polymerase, Reserve transcriptase}
- b} Vectors:- Cloning vectors [plasmid -pBR322,Bacteriophage-Lambda phage,Viruses-SV40, Cosmids vectors] and Expression vectors {Shuttle vector }
- Techniques: a] Southern, Northern and Western blotting b] PCR (Polymerase chain reaction)
- DNA Sequencing(Sanger)

UNIT-IV

- Gene cloning
- Linking of desired gene with vector DNA
- Introduction of recombinant DNA into host Cell
- Identification of recombinant DNA
- c-DNA libraries and Genomic libraries
- Transgenesis and Transgenic animals [Transgenic cattle, sheep, pig and fish]
- Animal cloning and cloned animals [Dolly sheep]
- DNA fingerprinting

- 1. Molecular Biology –David Friefelder –Narosa Publishing House, New Delhi.
- 2. Cell Biology, Genetics, Molecular Biology, Evolution & Ecology Verma, Agarwal S. Chand& Co.
- 3. Molecular & Cell Biology –Bhamrah –Anmol Publ. Pvt. Ltd., New Delhi.
- 4. Molecular Biology of the Cell Alberts, Bray, Lewis, Raff, Roberts, Watson Garland Publishers, New York.
- 5.Molecular Biology of the gene –J. D. Watson, NH Hopkins, Roberts, Stertz, Weiner-Freeman. 6.Concepts in Biotechnology 6.Editors-Balasubramanian, Bryee, Dharmalingam, Green, Jayraman Sangam Books.
- 7. Molecular Biology of the Gene Watson, Hopkins, Roberts, Steitz, Weiner Benjamin Cummings Publishing Co.
- 8. Molecular Cell Biology Baltimore, Zipursky, Matsudaria, Darnel W. H. Freeman & Co., New York.
- 9. Outlines of Biochemistry Conn & Stumpf.
- 10.Principles of Biochemistry White, Handler, Smith McGraw Hill Publ. 11.Cell & Molecular Biology Phillip Sheller Wiley Publ.
- 11. Molecular Biology Robert F. Wiver

B. Sc. II ZOOLOGY (SEM-IV)

Course Code: U-ZOO-382

Lab Course-VI: Biochemistry (CCZP-IV-A)

Credits: 02 Marks: 50 Lectures: 45

Course Objectives:

- They should standardize their solutions individually.
- To make the students to understand the skills necessary to analyze chemicals and chemical reactions quantitatively and qualitatively;
- To make the students to understand the skills necessary to verify hypothesis.
- To make the students to understand skills of titrimertic, Colorimetric lodometric analysis of vitamins, amino acid.

Course Outcome:

- Learners would understand the qualitative and quantitative analytical skill of biomolecules.
- Learners would understand skills of titrimertic, colorimetric, lodometric analysis of biomolecules.
- 1. Estimation of amino acids by formal titration.
- 2. Estimation of ascorbic acid by titrimertic method using 2, 6-dichlorophenol indophenol.
- 3. Estimation of Anioxidant by, 2-diphenyl-1-picrylhydrazyl (DPPH) radical scavenging assay
- 4. Estimation of reducing sugar from biological fluids by Benedict's titrimertic method.
- 5. Qualitative estimation of protein, Lipid and carbohydrates
- 6... Estimation of Protein by Biuret method/Bradford method/Lawry.
- 7. Estimation of amino acids by ninhydrin method
- 8. Isolation of glycogen from liver source and its estimation by anthrone method
- 9. Estimation of Lipids by Vanillin
- 10. Extraction of DNA by Phenol and Chloroform Method
- 11. Effect of temperature on enzyme action
- 12. Effect of pH on enzyme action
- 13. Effect of temperature on enzyme action
- 14. Effect of temperature on enzyme action

B.Sc. II ZOOLOGY (SEM-IV)

Course Code: U-ZOO-382

Lab Course-VII: Molecular Biology and Genetic Engineering (CCZP-IV-B)

Credits: 02 Marks: 50 Lectures: 45

Course Outcomes:

- To make the students to understand knowledge restriction digestion and r-DNA.
- To understand the applications of Molecular Biology and Genetic Engineering
- To have basic knowledge of cloning and Gene Expression, gene transfer, Blotting techniques, Equipments used in genetic etc.

Course Outcomes:

- Learners will able to understand the blotting technique.
- Learner will be able to use the agarose gel electrophoresis for DNA molecular size determination and will be able to estimate the DNA and RNA.
- Learner will able to understand the vectors in cloning techniques and rDNA technology.

Practicals:

- 1. Extraction of genomic DNA from Blood.
- 2. Estimation of Protein by Polyacrylamide Gel Electrophoresis
- 3. Estimation of DNA by Agarose gel electrophoresis
- 4. Estimation o RNA by Agarose gel electrophoresis
- 5 Demonstration DNA amplification by RT-PCR
- 6. Digestion of DNA with Restriction Enzymes
- 7. Demonstration Western Blotting
- 8. Demonstration of Southern / western blotting.
- 9. Estimation of DNA by DPA
- 10. Estimation of RNA by Orcinol reagent.
- 11 Extraction of proteins from tissues

B. Sc. II ZOOLOGY (SEM-IV)

Course Code: U-ZOO-382 Course: Microtechnique (SECZ-II)

Credits: 02 Marks: 50 Lectures: 45

Learning Objective:

- To understand collection of tissues and fixation techniques
- To understand Methods of block preparation
- To understand Sectioning and staining techniques.

Learning Outcomes:

After completion of this course students should be able to:

- Perform preparation of paraffin block of different tissues
- Perform fixation of tissues
- Perform sectioning and staining.

UNIT - I

- Introduction Definition of Histotechnology
- Methods of examination of tissues and cells, Collection and labeling of specimens, Methods of preparation and examination of tissues (fresh and fixed tissue)

UNIT-II

- Fixation of tissue Definition, Criteria for an ideal fixative, types (Simple and Compound), Properties of Simple and Compounds fixatives (Microanatomical, cytological and histochemical)
- Practical Isolation and collection of tissue, fixing and block preparation.

UNIT - III

- Tissue processing Manual and automatic tissue processing, Different embedding media, Steps of tissue processing (Dehydration, Clearing, and Impregnation).
- Embedding- Methods of Embedding, Embedding medium, names of medium and moulds, Automatic Tissue Processes (Structure and Working, Advantages and Disadvantages).
- Practical Tissue processing of prepared blocks.

UNIT - IV

- Section Cutting Types of Microtome, Rotary Microtome -Parts and their functions, Microtome Knives- Types, Care and Maintenance Techniques of sharpening; Technique of Section Cutting, Preparation of Adhesive Mixture, Mounting.
- Staining Definition and Significance of Staining, Stain and Staining Types, Theory of Staining, Methods of Staining.
- Practical Section Cutting, fixing, alcohol grading, staining and preparation of permanent slide.

REFERENCE BOOKS:

Histochemical Techniques – J. D. Bancroft.

Handbook of Histopathological and Histochemical Techniques - C.F.A. Culling.

Histological and Histochemical Methods 4th Edition – John Kier