

Special Issue (NSAZ-2022)







www.sciencejournal.in

Science and Engineering Research Board (SERB) Sponsored
National Symposium
On

Applied Zoology, Profitable Animal Production, and Health: Current Status and Future Progress (NSAZ-2022)

Organized by

Department of Zoology and Fishery Science, Rajarshi Shahu Mahavidyalaya (Autonomous), Latur- 413531, Maharashtra

On

23rd & 24th September- 2022

Volume 11, Issue 1(2022), ISSN: 2319-474X (p); 2319-4758 (e) © 2019 DAMA International. All rights reserved



National Symposium on

Applied Zoology, Profitable Animal Production, and Health: Current Status and Future Progress

(NSAZ-2022)

Organized by

Department of Zoology and Fishery Science, Rajarshi Shahu Mahavidyalaya (Autonomous),

Latur- 413531, Maharashtra

23rd & 24th September- 2022

(Conference Special Issue)

Copyright: © The Research Work as a Theory with Other Contents, Images, Tables, Charts in

Full Papers Are subject To Copyright Taken by Shiv Chhatrapati Shikshan Sanstha's Rajarshi

Shahu Mahavidyalaya (Autonomous), Latur Department of Zoology and Fishery Science (M.S.)

India, And © 2019 DAMA International. All Rights Reserved Executive Editors, Editors, Co-

Editors And Authors Of This Conference Issue.

Disclaimer: The author/authors are solely responsible for the content of the papers compiled

in this conference special issue. The publisher or editors does not take any responsibility for

the same in any manner. No part of this publication may be reproduced or transmitted in any

form by any means, electronic or mechanical, including photocopy, recording, or any

information storage and retrieval system, without permission in writing from the copyright

owner.

Published By:

Publisher: DAMA International

E-ISSN: 2319-4758 Print ISSN: 2319-474X

URL: www.sciencejournal.in

Chief Editor:Dr Laxmikant B. Dama Contact email: trendsres@gmail.com

Address: 15 B. Vijaynager, Z.P. Colony, Bijapur Road, Solapur (M.S.), India.

Country: India

Volume 11, **Issue** 1(2022)

ISSN: 2319–474X (p); 2319–4758 (e)

© 2019 DAMA International. All rights reserved

Index

	Index	
Sr.No	Papers / Article Title – Name of Authors	Page
		Number
01	Isolation, Identification and Biochemical Characterization of Probiotic Bacteria	1-6
	Isolated from Fresh Water Fishes of Gharani River	
	Madhuri Y. Bhande, Dnyaneshwar S. Rathod, Datta A.Nalle	
02	Assessment of antibacterial potential of formulated ointment containing medicinal	7-12
	plants against some fish bacterial pathogens.	
	Shivaji G Jetithor , Datta A.Nalle	
03	Rediscription of a species Senga paithanensis, Kadam et.al., 1981 from fresh	13-19
	water fish, Mastacembelus armatus (Lacepede, 1800) from Beed district (M.S.)	
	India.	
	Asha Bidkar , Amol Thosa and K.S. Raut	
04	Monthly Comparative Study of Physico-Chemical Parameters Of Siddheshwar	20-31
"	Reservoir of District Hingoli, Maharashtra, India.	20 61
	Priyanka Patode, B.S.Salve	
05	Monthly Comparative Study of Zooplankton Diversity Of Siddheshwar Reservoir	32-37
	of District Hingoli, Maharashtra, India.	32 37
	Priyanka Patode B.S.Salve	
06	Microbial Quality Assessment of Frozen fish and fish processing materials from	38-44
00		30-44
	Latur city market.	
07	Mushtakh Hashmi, Datta A. Nalle	45.45
07	Turbidity, TDS and Transparency in Water Bodies of Ekrukh Dam from Tale	45-47
	Hipparga Taluka South Solapur (M.S.) India	
	Shashikala Laxman Bhalkare	40 ==
08	A Review on Significance of Nutritional Value of Fish for Human Health.	48-55
	Eknath Pawade , Hanumant Jagtap	
09	Socioeconomic status of fishermen communities around Niwali reservoir in	56-67
	Parbhani District, (M.S.), India.	
	Mr. Gajanan. S. Sargar1,Mr. Ashish .S. Hasekar 2, Dr. B. G. Thakare.	
10	Isolation of Chitinolytic Pseudomonasspecies from the shrimp shell waste	68-74
	PB Pawar, *DV Vedpathak , *SM Inchure	
11	Comparative Study Of Major Carps DNA-RNA Ratio At Same Acceptance In	75-80
	Relation To Nutritional Condition And Their Growth Rate In Latur, Dist Latur	
	[M.S.], India	
	Raut K.S., K.D.Savant., Nagime P.S., Mahamuni P.B , Mali.P.P	
12	Effects of the amino acids rich dietary feed supplement on the. Protease activity	81-86
	of Wallago attu subsequent to Stress condition	
	Datta Ashok Nalle, Madhuri Y. Bhande, Pratiksha Patil	
13	Comparative Physico-Chemical Profile Of Sukhana River In Aurangabad,	87-100
	(M.S.) India	
	'Sonawane S.D. And Shaikh J.D.'	
14	Study of water quality of Darphal (gawadi) Lake, Solapur, Maharashtra.	101-104
	Sujit D. Pawar, Shahaji S. Chandanshive, Smita K.Shimple	
15	Synthesis and antimicrobial activities of New 3-(chloromethyl)-2-(piperidin-1-	105-115
	yl)quinoline Derivatives.	
	Kalimoddin I. Momin1, Rajkumar U. Pokalwar	
16	A case study of Magur (Clarias gariepinus), the fish banned in India	116-129
10	Phadke S.V.	110 127
	1 Haune 5. V.	

17	Essential Composition and Analytical Methods of Honey.	130-134
	Yeshwant Patne ,Abhinay Surwase, Dhanshree M. Jagtap	
18	Impact of Dimethoate Toxicity with Special Reference To Histopathological	135-142
	Alteration In Intestine of The Freshwater Fish Rasbora Daniconius.	
	Dnyaneshwar S. Rathod, Milindkumar V. Lokhande	
19	Altered quality aspects of deteriorated seeds of peanuts (Arachishypogaea L.)	143-151
	With response to ageing	
	K.D.Savant*, Raut K.S**	
20	Effect of Natural Coagulant on Cham Cham Prepared By Using Goat Milk	152-164
	B.D. Landge*, R.B. Yedatkar**, M.D. Rathod***	
21	IMPACT OF ANIMAL MORTALITY IN INDIA	165-168
	*SM Dapkekar, **RB Yedatkar, ***VV Lute	





Microbial Quality Assessment of Frozen fish and fish processing materials from Latur city market.

Mushtakh Hashmi, Datta A. Nalle

Department of zoology and fishery science, Rajarshi Shahu Mahavidyalaya, [Autonomous] Latur 413512 Maharashtra.

Email: <u>iprometheous007@gmail.com</u>

ABSTRACT

The present study aims at the microbiological analysis of export oriented frozen fishes, and fish processing water and ice from a view of public health safety and international trade. Microbiological analysis includes the determination of total viable aerobic count by standard plate count method and enumeration of total coliforms and fecal coliforms by most probable number method. The presence of specific fish pathogens such as *Salmonella* spp. and *Vibrio Cholerae* were also investigated. The TVAC of all the samples was estimated below cfu/g whereas the total coliforms and fecal coliforms count were found below 100 MPN/g and 10 MPN/g, respectively, which meet the acceptable limit specified by International Commission of Microbiological Specification for Food. The microbiological analysis of water and ice also complies with the specifications having cfu/mL, and total coliforms and fecal coliforms count were below the limit detection of the MPN method. Specific fish pathogens such as *Salmonella* sp. and *V. Cholerae* were found absent in all the samples under the investigation. From this study, it can be concluded that the investigated frozen fishes were eligible for export purpose and also safe for human consumption.

Keywords: microbial quality, frozen fish, sea food, cuttle fish, V. Cholera.

Introduction

Fish and fishery products are not only nutritionally important but also important in global trade as foreign exchange earner for a number of countries in the world [1]. Fisheries and aquaculture sectors have become the second most important contributors in export earnings of India, providing about 3.74% in national GDP, 2.7% in export earnings, and 22.23% in agriculture sector [2]. Due to wide range of global market including USA, UK, Japan, Belgium, Netherlands, Thailand, Germany, China, France, Canada, Spain, and Italy, the export of frozen fish, dry fish, and salted and dehydrated fish is increasing day by day from India. Fish are of great concern for export earnings because of their higher nutritive value such as high protein content, with little or no carbohydrate and fat value. But fish may be contaminated at various stages of transport, handling, and processing. This contamination may be related to the raw materials, personnel, and processing tools such as forklifts through leakage, insect, and pest harborage. Additionally, seafood can become contaminated during storage and processing [3, 4]. Contamination may be caused by foodborne pathogens which are naturally present in aquatic environments, such as Vibrio spp., or derived from sewage contaminated water such as Salmonella spp. [5]. Consumption of these





contaminated fish may cause infection or intoxication to the consumers. Vibrio cholerae is responsible for the third-highest number of shellfish-related illnesses, after noncholera Vibrio spp. and Norwalk viruses [6]. Toxigenic Ol (epidemic biotype) infections are associated with profuse, watery diarrhea whereas nontoxigenic, non-Ol biotype (except O139) infections result in septicaemia and mild gastroenteritis. In contrast to Vibrio spp., the incidence of Salmonella infections due to seafood consumption is still low compared with salmonellosis associated with other foods. However, detection of Salmonella spp. in seafood cannot be skipped as it is responsible for most of the foodborne diseases or gastroenteritis characterized by diarrhea, abdominal cramp, vomiting, nausea, and fever. According to Centers for Disease Control and Prevention, Salmonella is the leading cause of bacterial foodborne illness causing approximately 1.4 million nontyphoidal illnesses, 15,000 hospitalizations, and 400 deaths in the USA annually [7]. Water and ice quality is also an important factor for good quality fish, because water and ice used for fish processing may contaminate the whole processing plant. EU advised India Government to implement the Hazard Analysis Critical Control Point (HACCP) in the processing of frozen fishes [8]. So it is important to find out the quality of fish we consume as well as of the frozen fish which are exported. Therefore, the present study was carried out to investigate the microbiological quality of the marine frozen fishes for raising food safety concern and promoting international trade. This study also investigated the microbiological quality of water and ice, as these factors were intimately correlated with the fish processing and preservation.

Materials and Methods

Experimental Design:

Study Area

The district Latur (18.4088° N, 76.5604° E) this district has an historical background. The King 'Amoghvarsha' of Rashtrakutas developed the Latur city, originally the native place of the Rashtrakutas. The Rashtrakutas who succeeded the Chalukyas of Badami in 753 A.D called themselves the residents of Lattalut. The Latur geographical location shows that it situated southeastern part of the Maharashtra state. It surrounds Maharashtra Karnataka boundary. The entire district of Latur is situated on the Balaghat plateau, 540 to 638 meters from the mean sea level.

During the study period, total viable aerobic count, total coliforms and fecal coliforms counts, and presence of pathogenic organisms (namely, Salmonella spp. and Vibrio cholerae) of public health significance from the frozen fishes (storage temperature –20°C) and water and ice which were used during the processing of samples were investigated. All the frozen fishes were gutted and organoleptically good enough to carry out further bacteriological analysis. Sampling was done each year at three months' interval, during study periods, triplicate samples for each fish species as well as for ice and water samples were analyzed independently.

Chemicals and Media





Pure and analytical grade chemicals purchased from BDH were used throughout the study including media preparation. All the media and media ingredients such as beef extract and peptone that are used throughout the study. For the enumeration of coliforms and fecal coliforms. Lauryl

that are used throughout the study. For the enumeration of coliforms and fecal coliforms, Lauryl Tryptose Broth (LTB) and 2% Brilliant Green Bile Broth (BGLBB) were used, respectively. Bismuth Sulfite Agar (BSA) and Xylose Lysine Deoxycholate (XLD) agar were used for the detection of *Salmonella* spp. whereas Thiosulfate Citrate Bile Salt (TCBS) agar and Cellobiose, Polymyxin, and Colistin (CPC) were used for the detection of *V. cholerae*.

1.1 Fish Samples Preparation

All glassware was sterilized (121°C, 15 psi, 20 minutes) before use. Triplicate fish samples (each about 25 g) of each fish type were measured separately in an analytical balance in aseptic condition and then dissolved into about 225 mL buffered peptone water (BPW) and blended for (30–60) seconds in a sterilized blender machine [8]. Each fish sample was blended and homogenized separately.

1.2 Water and Ice Collection

The water (namely, WS1, WS2, and WS3) and ice (namely, IS1, IS2, and IS3) were collected in 1-liter sterilized container from different location of Latur fish market. The collected samples were preserved in the refrigerator (4°C), when analysis was delayed for more than 3 hours.

1.3 Enumeration of TVAC of Fish, Water, and Ice

Total viable aerobic bacteria of fish, water, and ice were enumerated by standard[9]. Plate count (SPC) method For the enumeration of TVAC, serial dilution of each sample was carried out.

After incubation, the plates having well spaced colonies (30–300) were used for counting and the colonies were counted by a colony counter, Total viable aerobic count per mL . multiplying the average number of colonies per plate by reciprocal of the dilution and expressed as colony forming units (cfu) per mL or per g of sample [10].

1.4 Enumeration of Total Coliforms of Fish

Most Probable Number (MPN) method is used for the quantitative estimation for coliform [11]. Serial dilution of the samples was prepared as described earlier. Nine test tubes containing about 9 mL Lauryl Tryptose Broth

Enumeration of Fecal Coliforms (Presumptive E. COLI Test) of Fish





About one loopful from each gas positive LTB was inoculated into test tube of sterilized BGLBB and a test tube of sterilized 10 mL Tryptone Broth and then incubated at for 48 hours. After incubation, gas production was recorded and 2-3 drops of Kovac's reagent were added to each of the positive tubes for total fecal coliforms number (E. coli) per gram [12]. Enumeration of Total Coliforms and Fecal Coliforms in Water and Ice About 50 mL of water was inoculated to 50 mL of sterilized LTB (double strength) in one mega test tube whereas about 10 mL of water was inoculated in five test tubes containing 5 mL of sterile LTB (double strength) Coliforms of ice samples were also enumerated similarl Detection of Salmonella spp. About 25 g samples were dissolved in about 225 mL of sterilized buffered peptone water (BPW), blended, and incubated at 37°C for 16–20 hours. About 10 mL from the incubated BPW culture was selectively enriched into the 100 mL sterilized Selenite Cystine Broth and incubated again at 37°C for 24-48 hours. biochemical characteristics, namely, TSI (Triple Sugar Iron), Urease test, MR-VP test, Oxidase test, Citrate test, fermentation of carbohydrates (Glucose, Sucrose, Arabinose, Mannose, Mannitol, and Inositol), and Decarboxylase (Lysine, Arginine, and Ornithine) following the taxonomic guides of Bergey's Manual of Determinative Bacteriology, 8th ed. [14]. All cultures giving biochemical reactions were confirmed by agglutination test with Salmonella polyvalent (O) somatic antisera [15].

Detection of Vibrio

Vibrio spp was detected following the procedure as described by Kaysner and Angelo [16] (Glucose, Sucrose, Arabinose, Mannose, Mannitol, and Inositol), and Decarboxylase test (Lysine, Arginine, and Ornithine) according to the taxonomic guides of *Bergey's Manual of Determinative Bacteriology*, 8th ed. [14]. Finally the *V. cholerae* were confirmed by agglutination test using polyvalent *V. cholerae* (O) antiserum [15].

Results

Fish and seafoods hold an important position as a food component for a large section of world population [17].. So, maintenance of appropriate quality of the products is regarded as vital for achieving desired success in the global trade of this product. Jew fish (*A. hololepidotus*), Queen fish (*S. commersonnianus*), Tongue Sole fish (*C. broadhursti*), Ribbon fish (*L. savala*), and Cuttle fish (*S. officinalis*) are the most commonly exported marine fishes from India.

The maximum microbiological limit for the TVAC which separates the good quality products from bad quality is 5×10^5 cfu/g [18].

The TVAC of the studied samples ranged from 2.8×105 to 4.9×105 cfu/g which was below the maximum acceptable limit. So all the samples of each type of the fish meet the acceptable limit specified by ICMSF which points out the good quality of the frozen fishes showed the TVAC of all





frozen fish samples from which it was observed that the density of total aerobic bacteria detected in the Tongue Sole fish samples was comparatively higher than all of the fishes whereas the lowest bacterial count was observed in the samples of Jew fish. Loads of bacteria in fish samples decreased gradually over time in all of the fishes. This may be due to the aseptic processing and handling, proper sampling, trained personnel, improved storage conditions, and increased awareness for preservation The acceptable limits of total coliforms (TC) and fecal coliforms (FC) for fresh and frozen fish are <100 MPN/g and <10 MPN/g, respectively [18]. The presence of TC is indicator of sewage contamination which may also occur during different processing steps such as transport and handling. Moreover, the contamination may also be caused by the water used for washing or icing [19]. The more accurate indicator of fecal contamination is fecal coliforms that is E. coli [20]. The lower number of coliforms can be beneficial for pointing out the effectiveness of safety procedures during processing and handling [21]. In the present study, the total coliforms count ranged from 5 MPN/g to 28 MPN/g and fecal coliforms count was from 3 MPN/g to 8.3 MPN/g. Figures 2 and 3 showed the highest number of coliforms and fecal coliforms bacteria in Tongue Sole fish whereas the lowest count was observed in the samples of Jew fish, respectively. Our study revealed that all samples were within the recommended limits which indicated that the samples were collected from pollution-free water and also the food processors and handlers maintained aseptic conditions throughout the processing. Water and ice are the most important factor for the processing of exported fish. These two factors contribute to determining and maintaining of the standard quality of the frozen fishes. Figures 4 and 5 showed the TVAC of water and ice samples, respectively, over the study period. It was found that TVAC of water samples and ice samples ranged from 3 to 18 cfu/mL. Significant reduction of TVAC for both water and ice samples was observed over the time period. The total coliforms and fecal coliforms count were found absent for both samples. Hence, our study revealed that all the tested samples complied with the recommended limit specified by ICMSF, that is, TVAC having <20 cfu/mL, and coliforms and fecal coliforms count were below the limit detection of the MPN method. This may be due to the advanced and improved facilities for water and ice purification, treatment, and handling. Seafood infections are caused by variety of bacteria, viruses, and parasites. According to Centers for Disease Prevention and Control (CDC), during 1973 to 2006, 188 outbreaks of seafood-associated infections, causing 4,020 illnesses, 161 hospitalizations, and 11 deaths, were reported to the Foodborne Disease Outbreak Surveillance System. Most of these seafood-associated outbreaks (143 (76.1%)) were due to a bacterial agent; 40 (21.3%) outbreaks had a viral etiology; and 5 (2.6%) had a parasitic cause. According to the report, Vibrio were the most commonly reported cause of seafood-associated outbreaks where toxigenic V. cholerae caused 3 outbreaks and 10 illnesses without deaths and non-O1, non-O139 V. cholerae caused 4 outbreaks and 12 illnesses without deaths, whereas Salmonella was responsible for 18 outbreaks, 374 illnesses, and 28 hospitalizations durin g the study period





Recently CDC reported that about 62 people were infected with *Salmonella* Paratyphi B variant L (+) tartrate (+) (formerly known as *Salmonella* Java) from 11 states of USA related to the consumption of frozen raw tuna. The infection was characterized by diarrhea, fever, and abdominal cramps after 12–72 hours' exposure without paratyphoid fever, enteric fever, or typhoid fever [23].

Conclusion:

Although seafood is part of a healthful diet, its consumption is not out of risk. Worldwide continued outbreaks of seafood-associated infections have rendered the existing control strategies questionable. An understanding of the etiologic agents, seafood commodities associated with illness, and mechanisms of contamination that are amenable to control is thus necessary for the prevention of seafood-associated infection outbreaks

Coordinated efforts from government sector and private industry together with federal agencies are urgently needed in this context. There is a need for routine surveillance systems using pathogen-specific techniques to avoid any future outbreak

However, the current study revealed that microbiological quality of the investigated frozen fishes and fish processing materials (ice and water) was within the specified limit of ICMSF. So it can be concluded that these fishes were processed with properly treated pathogen-free water and ice and, finally, maintained at good storage condition. Hence, the investigated frozen fishes were qualified enough for export as well as human consumption from bacteriological point of view. The presence of viruses, parasites, viable but nonculturable (VBNC) state of the pathogenic bacteria, and biochemical parameters such as histamine risk might be a problem in frozen fish products which is the limitations of this study. Beyond ICMSF, in order to comply with more stringent indigenous quality standards of the exporting countries, these quality parameters must be taken into consideration.

References

- 1. S. O. Yagoub and T. M. Ahmed, "Pathogenic Microorganisms in fresh water samples collected from Khartoum central market," *Sudan Journal of Veterinary Science and Animal Husbandry*, vol. 43, no. 1-2, pp. 32–37, 2003.
- 2. F. L. Bryan, "Epidemiology of foodborne diseases transmitted by fish, shellfish and marine crustaceans in the United States, 1970–1978," *Journal of Food Protection*, vol. 43, pp. 859–876, 1980.





- www.sciencejournal.in
- 3. E. J. Gangarosa, A. L. Bisno, E. R. Eichner et al., "Epidemic of febrile gastroenteritis due to Salmonella java traced to smoked whitefish," *American Journal of Public Health*, vol. 58, no. 1, pp. 114–121, 1968.
- 4. K. Gnanambal and J. Patterson, "Biochemical and microbiological quality of frozen fishes available in Tuticorin supermarkets," *Fishery Technology*, vol. 42, no. 1, pp. 83–84, 2005.
- 5. R. J. Wittman and G. J. Flick, "Microbial contamination of shellfish: prevalence, risk to human health, and control strategies," *Annual Review of Public Health*, vol. 16, pp. 123–140, 1995.
- Centers for Disease Control and Prevention (CDC), "Preliminary foodnet data on the incidence of infection with pathogens transmitted commonly through food," *Morbidity and Mortality Weekly Report*, vol. 59, no. 14, pp. 418–422, 2010.
- 7. W. H. Andrews and T. S. Hammack, "Food sampling and preparation of sample homogenate," in *United States Food and Drug Administration (US FDA) Bacteriological Analytical Manual*, chapter 1, United States Food and Drug Administration, Silver Spring, Md, USA.
- 8. 2001 L. J. Maturin and J. T. Peeler, "Aerobic plate count," in *Bacteriological Analytical Manual*, chapter3, United States Food and Drug Administration (USFDA) 2001.C. H. Collins and M. P. Lyne, Microbiological Methods, Butterworth, London, UK, 5th edition, 1984.
- 9. P. Feng, D. W. Stephen, and A. G. Michael, "Enumeration of Escherichia coli and the coliform bacteria," in Bacteriological Analytical Manual, chapter 4, United States Food and Drug Administration (USFDA), 2002J. L. Oblinger and J. A. Koburger, "Understanding and teaching the most probable number technique," Journal of Milk and Food Technology, vol. 38, pp. 540–545, 1975.
- 10. W. H. Andrews and T. S. Hammack, "Salmonella," in United States Food and Drug Administration (US FDA) Bacteriological Analytical Manual, chapter 5, United States Food and Drug Administration, Silver Spring, Md, USA, 2007.
- 11. R. E. Buchanan and N. E. Gibbons, *Bergeys Manual of Determinative Bacteriology*, The Williams and Wilkins Company, Baltimore, Md, USA, 8th edition, 1974.
- 12. P. K. Surendran, N. Thampuran, and K. Gopakumar, "Microbial profile of cultured fishes 1 and prawn viz a viz their spoilage and contamination," in *Proceedings of the 9th Session of the Indo-Pacific 2 Fishery Commission Working Party on Fish Technology and Marketing*, D. James, Ed., vol. 3, pp. 1–12, FAO, Rome,
- 13. C. A. Kaysner and D. J. Angelo, "Vibrio," in United States Food and Drug Administration (US FDA)
 - *Bacteriological Analytical Manual*, chapter 9,United States Food and Drug Administration, Silver Spring, Md, USA, 2004.





- 14. W. M. K. Bakr, W. A. Hazzah, and A. F. Abaza, "Detection of Salmonella and Vibrio species in some seafood in Alexandria," Journal of American Science, vol. 7, no. 9, pp. 663–668, 2011.
- 15. ICMSF (International Commission of Microbiological Specification for Food), Microorganisms in Food 2. Sampling for Microbiological Analysis: Principles and Specific Applications, University of Toronto Press, Toronto, Canada, 2nd edition, 1986.
- 16. C. E. Boyd, Water Quality in Ponds for Aquaculture, Alabama Agricultural Experiment Station, Auburn University, Auburn, Ala, USA, 1990.
- 17. V. Suvanich, D. L. Marshall, and M. L. Jahncke, "Microbiological and color quality changes of channel catfish frame mince during chilled and frozen storage," Journal of Food Science, vol. 65, no. 1, pp. 151–154, 2000.
- 18. N. Elhadi, S. Radu, C.-H. Chen, and M. Nishibuchi, "Prevalence of potentially pathogenic *Vibrio* species in the seafood marketed in Malaysia," *Journal of Food Protection*, vol. 67, no. 7, pp. 1469–1475, 2004.
- 19. M. Iwamoto, T. Ayers, B. E. Mahon, and D. L. Swerdlow, "Epidemiology of seafood-associated infections in the United States," *Clinical Microbiology Reviews*, vol. 23, no. 2, pp. 399–411, 2010.
- 20. Centers for Disease Prevention and Control (CDC), "Multistate outbreak of *Salmonella* paratyphi B variant L(+) tartrate(+) infections linked to frozen raw tuna," July 2015.